

The *mPlrp2* and *mClps* genes are involved in the hydrolysis of retinyl esters in the mouse liver

Wenqiang Pang ^{#1}, Ying Zhang ^{#1}, Shiming Wang¹, Aiqun Jia¹, Wei Dong¹, Chun Cai¹,
Zichun Hua² and Jianfa Zhang^{1*}

From¹Center for Molecular Metabolism, Nanjing University of Science & Technology, Nanjing, 210094, China; ²The State Key Laboratory of Pharmaceutical Biotechnology, Nanjing University, Nanjing, 210090, China

These authors contributed equally to this work.

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*Address correspondence to: Jianfa Zhang, Ph.D, B508, No. 364 building, 200 Xiaolinwei Street, Nanjing, 210094, China. Fax: 86-25-84318533; Email: jfzhang@mail.njust.edu.cn

Abstract

Retinyl esters are the major chemical forms of vitamin A stored in the liver, which can be delivered to peripheral tissues for conversion into biologically active forms. The function and regulation of the hepatic genes that are potentially involved in catalyzing the hydrolysis of retinyl esters remain unclear. Here we show that two lipid hydrolytic genes, pancreatic related protein 2 (*mPlrp2*) and procolipase (*mClps*), expressed specifically in the mouse pancreas, are associated with the ratio of S-adenosylmethionine (AdoMet) to S-adenosylhomocysteine (AdoHcy). Light illumination deficiency or administration of 5'-adenosine monophosphate (5' AMP) elevated the ratio of AdoMet to AdoHcy and induced the expression in the liver of *mPlrp2* and *mClps*, which was blocked by all-trans retinoic acid (ATRA). Mice fed a vitamin A free diet exhibited increased activation of hepatic *mPlrp2* and *mClps* expression, which was associated with increased methylation of histone H3K4 residues located near the *mPlrp2* and *mClps* promoters. Inhibition of hepatic *mPlrp2* and *mClps* expression by a methylase inhibitor, methylthioadenosine (MTA), markedly decreased plasma retinol levels in these mice. The activated HSC-T6 cell line specifically expressed *mClps* and *mPlrp2*. Inhibition of *mClps* gene expressions by shRNA decreased hydrolysis of retinyl esters in the HSC-T6 cell line. These data suggest that the conditional expression of *mPlrp2* and *mClps* is involved in the hydrolysis of retinyl esters in the mouse liver.

Key words: AdoMet; AdoHcy; mClps; mPlrp2; Hepatic retinyl ester

Introduction

Vitamin A and its derivatives (retinoids) play an essential role in differentiation and development by their conversion to active form retinoic acid, which interacts with the nuclear receptors of the RXR and RAR families, functioning as hormone-activated factors in the regulation of the expression of multiple genes (1,2). Also, vitamin A mediates vision cycles through its metabolite, 11-cis retinaldehyde, which is the active chromophore in rhodopsin (3, 4). Most of the retinoids in the body are accounted for by the retinyl esters stored in the liver (5, 6). The formation and hydrolysis of retinyl esters are key reactions for maintaining a constant concentration of free retinol in circulation. It has been demonstrated that lecithin:retinol acyltransferase and acylCoA:retinol acyltransferase contribute to the storage of retinoids by the conversion of circulatory free retinol to retinyl esters (7, 8). The enzymes catalyzing the reverse reaction, designated retinyl ester hydrolase, promote the liberation of retinol from stored retinyl esters in the liver into general circulation. Mice fed Vitamin A-deficient diet for 6 weeks decreased the hepatic retinyl esters (9), and Vitamin A-deficient rats obviously impaired the liver uptake of retinol from blood (10). However, the identity, function and regulation of hepatic enzymes potentially involved in catalyzing the hydrolysis of retinyl esters remain unclear (11, 12).

The bile salt-activated carboxyl ester lipase, an enzyme that was found in the pancreas and liver cells of several mammalian species, had been thought to be the main hepatic retinyl ester hydrolase (13). Mice deficient in carboxyl ester lipase exhibit normal levels of intestinal and hepatic uptake of retinyl esters and maintain retinyl ester hydrolase activity in the liver and pancreas, suggesting that in the mouse liver and pancreas, there is a retinyl ester hydrolase distinct from carboxyl ester lipase (14). Pancreatic triglyceride lipase (PTL) and its partner, colipase, have been demonstrated to be major pancreatic retinyl ester hydrolases (REH) in mice. Like classical PTL, the pancreatic lipase related protein 2 (PLRP2), with strong nucleotide and amino acid sequence homology to PTL, also cleaves triglycerides and retinyl esters, but PLRP2 has broader substrate specificity (15, 16). Although PLRP2 does not absolutely require colipase, colipase may increase PLRP2 protein activity (17). In suckling mice, PLRP2 protein is the major colipase-dependent pancreatic lipase (18).

The murine genes encoding PLRP2 (*mPlrp2*) and colipase (*mClps*) are expressed specifically in the pancreas to hydrolyze fat and retinyl esters in the diet (19). Our recent studies demonstrate that a reduction in light illumination results in upregulated expression of *mPlrp2* and *mClps* in the mouse liver (20), and that the light-modulated all-trans retinoic acid (ATRA) signal plays a key role in the regulation of the expression of these genes (21). In this study, we characterized the molecular mechanism by which *mPlrp2* and *mClps* are expressed specifically in the pancreas by assessing the difference in the ratio of AdoMet to AdoHcy between the pancreas and the liver. A temporary, light-modulated change in the ratio of AdoMet to AdoHcy regulates hepatic *mPlrp2* and *mClps* gene expression in mice. Using mice fed a vitamin A free diet, hepatic *mPlrp2* and *mClps* gene expression was found to be directly related to the level of retinol in the plasma. The results of the studies strongly suggest that hepatic *mPlrp2* and *mClps* gene expression is associated with the hydrolysis of retinyl esters in the mouse liver.

Experimental Procedures

Animals. 6-8 weekold female C57BL/6 mice were used in this work. Mice were maintained in 12 /12-h light/dark (LD) cycles with lights on at 7:00 am and off at 7:00 pm and allowed free access to food and water. For VA-free diet experiment, mice were placed on a vitamin A-free diet for 1 week, and the control animals were given normal chow. The vitamin A-free diet (Laboratory, Nanjing, China) for 100 g food by dry weight was composed of vitamin-free casein (18%); sucrose plus maize starch (68.4%); cellulose (2%); hydrogenated coconut oil (4.6%); salt (4.8%); yeast (2%); and a vitamin integration lacking Vitamin A (0.2%). A control group of mice was fed using standard laboratory chow. For consistent observations, all samples were collected at 7:00 pm, unless otherwise indicated. All animal care and use procedures were in accordance with the guidelines of the Institutional Animal Care and Use Committee at NJUST.

Light illumination analysis. For light quality experiments, animals were transferred to closed chambers illuminated by light-emitting diode (LED) illuminators (PARA, Nanjing, China); the intensity of colored light was matched to the intensity of the incident white light source by adjusting the distance between the light and the cage covers (about 250 lux). Colored light was generated by the following LED (Burbank, CA): blue (470 nm), green (528 nm), yellow (593 nm) and red (625 nm). Mice were maintained in colored light/dark (12/12-h) or constant darkness (DD) at the same schedule as the white cycle for 72 h. For light intensity experiments, animals were transferred from standard light (about 250 lux) to weak light (about 20 lux at the cage cover) from a white light lamp in 12/12-h light/dark cycles for 72 h. Samples were collected at the time points indicated in the figure legends. Light intensity was measured by a Laboratory Digital light Meter CT-1330B (Nanjing, China).

RNA isolation and RT-PCR Analysis. Total RNA was extracted from fresh liver samples with TriZol Reagent, and the remaining DNA was completely removed by RNase-free DNase treatment. Total RNA (50 ng) was reverse-transcribed with poly(dT)12–18 as a first-strand primer, according to the manufacturer's instructions. RT-PCR was performed and analyzed using an ABI 9700 Detection System. Quantitative real-time RT-PCR was performed and results were analyzed using an ABI 7300 Detection System utilizing SYBR Green dye. The following primers were designed: *Gapdh*: 5'-CATCCACTGGTGCTGCCAAGGCTGT-3' (forward), 5'-ACAAACCTGGTC CTCA GTGTAGCCCA-3' (reverse); *mPlrp2*: 5'-GGACCAAAGAA GCTGAGT GG-3' (forward), 5'-ATAGTGGG CCGGAA GAGATT-3' (reverse); *mClps*, 5'-CTTGTGT CCCTCCTTGCAGT-3' (forward), 5'-GAGCACTCGCTGTT TCC AT- 3' (reverse). Reaction conditions were as follows: cycle 1(1x): 50 °C for 2 min, cycle 2 (1x): 95 °C for 10 min, cycle 3 (35 x): 94 °C for 15 s, 58 °C for 30 s, 72 °C for 30 s. All primer sequences were located between more than two exons. Relative gene expression in comparison with *Gapdh* expression was calculated by the comparative CT method.

Western blotting. The protein samples were separated by 10% homogeneous SDS-PAGE before the proteins were transferred to nitrocellulose membranes. After blocking with milk in PBS-Tween (0.1%) buffer overnight at 4°C, the transferred membranes were incubated with PLRP2 antibody (Santa Cruz Biotechnology, USA) for 2 h at 26°C; followed by horseradish

peroxidase-conjugated secondary antibody (Santa Cruz Biotechnology) for 1 h at 26°C. The blotting was developed by using ECL detection reagent (Keygen, China) and visualized by autoradiography. HRP-conjugated monoclonal mouse β -Actin was introduced as a reference.

Determination of hepatic levels of AdoMet and AdoHcy. Extracts were separated and quantified by using reverse-phase HPLC (Waters 1525 system, Millipore Corp., Bedford, MA) analysis on a Partisphere bonded phase C18 (reverse phase) cartridge column, according to procedures previously described by She *et al* (22). The mobile phase contained 0.1 M sodium acetate, 5 mM heptanesulfonic acid adjusted to pH 4.50 with acetic acid, and 4.2% acetonitrile. The samples were eluted at room temperature with an invariable gradient at a flow-rate of 0.8 ml/min. Characteristic peak spectra and retention times compared to those of the standard were used to identify AdoMet and AdoHcy. Quantitation was based on peak areas. AdoMet and AdoHcy standards were purchased from Sigma (MO, USA).

Plasma retinol analysis by HPLC. Analysis of plasma retinol levels was performed by reverse phase HPLC according to the method described by Teerlink (23). Plasma was collected in EDTA-coated tubes, isolated and frozen in the dark at -70 °C until the determination of retinol concentrations. Retinoid extraction and analytical procedures were carried out in a darkened room under dim yellow light (15 w) to protect retinoids from exposure to light. The extraction procedure involved in single-phase fluid extraction consisted of adding 350 μ l of plasma to 50 μ l of 1.0 M sodium acetate buffer (pH 4.0). After mixing, 600 μ l of acetonitrile was added to the mixture, and the samples were immediately vortex-mixed and centrifuged for 5 min at 12000 g and 4°C, and 450 μ l of clear supernatant was decanted and diluted with 150 μ l of water. 400 μ l of this solution was injected for HPLC analysis. The analysis was performed on a Waters HPLC system (Waters 1525, USA) equipped with a vacuum degasser, quaternary pump, autosampler, column heater, and diode array detector. HPLC analysis was conducted on a 5 μ m octadecyl silane C₁₈ column (25 cm \times 4.6 mm internal diameter; CEL Associates, Pearland, TX). The mobile phase consisted of acetonitrile/ methanol / 2% ammonium acetate solution/ glacial acetic acid (79:2:16:3) with a flow rate of 1.0 ml/min. The temperature of the column was maintained at 30 °C. The diode array detector was set at a wavelength of 325 nm to monitor all compounds simultaneously. Characteristic peak spectra and retention times were compared with a retinol standard (Sigma, MO, USA) to identify retinol. Quantitation was based on peak areas.

Treating animals with 5'-AMP, MTA and ATRA. The indicated dosages of 5'-AMP, MTA and ATRA (Sigma, MO, USA) were administered by intraperitoneal injections in various groups of mice. After injection, mice were maintained for the desired period length and then sacrificed. Plasma or liver tissue was collected and extracted for analysis of AdoMet and AdoHcy ratios or retinol levels. Total RNA was isolated from liver tissues for RT-PCR analysis.

Chromatin immunoprecipitation (ChIP) assay. The ChIP assay was performed as previously described (24, 25), with some modifications for the liver. Briefly, 100 mg of liver tissue was cut into small pieces with a razor blade and cross-linked with 1% formaldehyde for 15 minutes. Crosslinking was stopped with 0.125 M glycine and samples were rinsed with 1 \times PBS containing 1 mM PMSF and disaggregated with a Dounce homogenizer. The nuclei were pelleted and resuspended in lysis buffer (1% SDS, 10 mM EDTA, 50 mM Tris-HCl at pH 8.1, 0.8 μ g/ ml of pepstatin A, 0.6 μ g/ml of leupeptin and 1 mM PMSF). The suspension

was sonicated using a sonicator (KS-130, Ninbo Kesheng, China) to generate DNA fragments averaging 400–500 bp in length and clarified by centrifugation. For immunoprecipitation, the supernatant was diluted 10 fold with dilution buffer (1% Triton X-100, 2 mM EDTA, 150 mM NaCl and 20 mM Tris-HCl at pH 8.1) and divided into fractions for control IgG and anti-trimethyl-H3K4 (Millipore). Protein A-Sepharose beads, pre-blocked with 300 µg/mL of sheared salmon sperm DNA and 0.1% BSA, were used to precipitate antibody-chromatin complexes. Beads were washed sequentially for 5 min each with TSE buffer with 150 and 500 mM NaCl, buffer III, and TE (pH 8.0). Immunocomplexes were eluted, the elution was heated to reverse the formaldehyde cross-linking and DNA was phenol:chloroform extracted. For PCR analysis of the ChIP samples, purified immunoprecipitates were dissolved in 20 µL of water.

Preparation of shRNA. The shRNAs targeting rat *mClps* mRNA were designed and synthesized by GenScript. Sequences used for the shRNAs in the experiments were: shRNA-1: 5'-TGCCAACATGACACCCTCTG-3', and shRNA-2: 5'-TGCAGTGTAAGAGCAGATGCT-3'. All of these sequences were shown by BLAST search not to share sequence homology with any rat mRNA. Negative control shRNAs was used to assess non-specific gene silencing effects. The mammalian expression vector pRNA-U6.1 was used to express shRNAs in HSC-T6. ShRNAs were introduced into HSC-T6 cells by GenEscortTM II, according to the manufacturer's protocol.

Cell Culture. HSC-T6 cells were cultured in DMEM (Invitrogen) containing 25 mM glucose, 10% (v/v) FBS and 100 U/mL penicillin/streptomycin at 37°C/5% CO₂. HSC-T6 cells were incubated for 12 h with palmitic acid (100 µM), after which the cells were treated with 5 µM retinol for an additional 12 h, and then shRNAs or the indicated dosages of MTA (4 µM and 12 µM) were introduced into the cells. Retinyl ester concentrations were determined 12 h later.

PLRP2 Immunohistochemistry. To detect PLRP2, slides of paraffin embedded liver sections were deparaffinized and rehydrated. Immunohistochemical staining for PLRP2 with a mouse antibody against PLRP2 (Santa Cruz Biotechnology, USA) was performed according to the manufacturer's protocol.

Statistical analysis. Data are presented as the mean ± the standard error of the mean (SEM). Statistical analysis was performed with a Student *t*-test. Significance was defined by *p* < 0.05.

Results

Weak light illumination activated the expression of *mPlrp2* and *mClps* in the liver. The lipase gene *mPlrp2* and its partner *mClps* are expressed specifically in the mouse pancreas in general conditions (26). Our previous work demonstrated that light exposure inhibits both *mPlrp2* and *mClps* gene expression and that darkness induce their expression in the mouse liver (20, 21). We hypothesized that a specific light wavelength or light intensity in illumination was a modulator for *mPlrp2* and *mClps* gene expression. Therefore, we carried out specific wavelength illumination studies to determine whether there was a difference in these genes' responses to light. Confirming previous observations, no hepatic *mPlrp2* and *mClps* gene expression was observed under light/dark (LD) cycles (Fig. 1A). Mice maintained in similar intensities of various wavelengths, including red light, yellow light, blue light and green light, failed to exhibit *mPlrp2* and *mClps* gene expression in the liver, indicating that the inhibition of hepatic *mPlrp2* and *mClps* expression by white light was not due to a specific wavelength (Fig. 1B). Next, we investigated whether absolute light deficiency (dark/dark, DD) was necessary to induce hepatic *mPlrp2* and *mClps* expression. RT-PCR analysis showed that light intensity below 20 lux activated *mPlrp2* and *mClps* gene expression in the liver (Fig. 1B), and no expression of these genes could be determined under high light intensity. These observations suggest that light intensity is a regulatory signal in modulating hepatic *mPlrp2* and *mClps* gene expression. Moreover, western blotting showed that DD induced PLRP2 protein expression, whereas light exposure for 2 h decreased the protein level and for 6h light illumination PLRP2 almost can't be detected (Supplementary Fig. 1).

Specific expression of *mPlrp2* and *mClps* was associated with the ratio of AdoMet/AdoHcy in the pancreas. Although light intensity plays a dominating role in inducing *mPlrp2* and *mClps* gene expression in the liver, light exposure or darkness failed to induce a significant difference in the expression pattern of either gene in the pancreas. In normal conditions for maintaining animals, the expression of the *mPlrp2* and *mClps* genes was tissue specific and restricted to the pancreas; both gene products functioned in hydrolyzing lipid from diet fat and retinyl esters (19). The conditional expression of *mPlrp2* and *mClps* in the liver by weak light illumination or darkness was reversible. In order to identify whether various states of energy metabolism in tissues could result in the specific expression of *mPlrp2* and *mClps*, we examined the difference between the pancreas and liver tissues. First, nucleotides were analyzed from extracts of both tissues. There was no significant difference in the levels of ATP, ADP and AMP between the pancreas and the liver, excluding the possibility that nucleotide metabolism results in the specific expression of *mPlrp2* and *mClps*. Then, we compared the ratio of AdoMet to AdoHcy, which represents the potential of methylation in tissue (27). HPLC profiles showed that the peak of AdoMet was much higher than that of AdoHcy in the pancreas (Fig. 2A), whereas liver tissues had no obvious differences in the peaks of AdoMet and AdoHcy (Fig. 2B). Next, we quantified the levels of the peaks obtained from the liver and pancreas (Fig. 2C, n=6). This analysis revealed that the ratio of AdoMet to AdoHcy was markedly higher in the pancreas, compared with that found in the liver.

Darkness or weak light illumination elevated the ratio of AdoMet to AdoHcy in livers. To

demonstrate that the ratio of AdoMet to AdoHcy was associated with hepatic *mPlrp2* and *mClps* expression, we measured AdoMet and AdoHcy in the mouse livers under darkness and weak light illumination. HPLC profiles showed peak AdoHcy declined and peak AdoMet rose in mouse livers in the DD cycle compared with the normal LD cycle (Fig. 3A). A similar change was observed in mice maintained under weak light illumination. Moreover, in DD animals, light exposure elevated the AdoHcy peak to a level comparable to that of LD animals (Fig. 3A). Quantitative analysis of peak levels revealed that the ratio of AdoMet to AdoHcy was markedly elevated in mouse livers under darkness compared with light illumination; the elevated level of the ratio in the liver was close to that found in the pancreas; and light exposure in DD mice for 4 h reversibly returned this ratio to the normal levels found in LD animals (Fig. 3B). Previous observations demonstrated that light exposure in DD mice inhibited *mPlrp2* and *mClps* expression in the livers (20, 21).

Administration of ATRA lowered the ratio of AdoMet to AdoHcy in DD livers, and administration of 5'-AMP increased the ratio of AdoMet to AdoHcy in LD livers. Our recent studies demonstrate that ATRA has a feedback inhibition to hepatic *mPlrp2* and *mClps*. The light-modulated ATRA signal regulated ecto-5'-nucleotidase (CD73) gene expression by the RARalpha receptor, affecting the circulatory levels of 5'-AMP, resulting in *mPlrp2* and *mClps* expression (21). Therefore, we carried out experiments to analyze the alteration of hepatic AdoMet and AdoHcy in ATRA and 5'-AMP treated animals. RT-PCR analysis showed that administration of 5'-AMP in LD animals activated *mPlrp2* and *mClps* gene expression in the liver (Fig. 4A), confirming a previous northern analysis; expectedly, LD livers indicated that the ratio of AdoMet to AdoHcy increased notably after 5'-AMP injection (Fig. 4B). Next, ATRA (2 nmol/g and 10 nmol/g)-treated mice exhibited reduced *mPlrp2* and *mClps* expression under DD conditions (Fig. 4C), and the ratio of AdoMet to AdoHcy declined in ATRA treated livers compared with control livers (Fig. 4D). These observations implied that the temporary changes of the ratio of AdoMet to AdoHcy in the livers were associated with hepatic *mPlrp2* and *mClps* expression.

Mice fed a vitamin A free diet activated hepatic *mPlrp2* and *mClps* expression, along with related increases in the methylation of histone H3K4 associated with the *mPlrp2* and *mClps* promoters. The gene products of *mPlrp2* and *mClps* have been demonstrated to function in the hydrolysis of dietary retinyl esters in the gastrointestinal organs (19), implying the possibility of similar hydrolysis activities for hepatic retinyl esters. To determine whether hepatic *mPlrp2* and *mClps* were related with the hepatic utilization of retinoids, mice were placed on a vitamin A free diet for 7 days. Surprisingly, hepatic *mPlrp2* and *mClps* mRNA were upregulated strongly in the livers of mice fed a vitamin A-free diet (Fig. 5A). Next, we detected the expression levels of other lipases in the liver, such as hepatic lipase, lipoprotein lipase and carboxylester lipase, and found no obvious reproducible changes in these lipases in mice fed the vitamin A-free diet compared with that observed in mice fed a normal diet (data not shown). A subsequent analysis of AdoMet and AdoHcy showed that the ratio of AdoMet to AdoHcy in the liver is elevated in these animals compared with mice fed a normal diet (Fig. 5B). These changes in AdoMet and AdoHcy levels resulted in an increase of total trimethylation of histone H3K4 related liver DNA in mice fed a vitamin A-free diet compared with control mice, and ChIP analysis indicated an increase in the binding of trimethylated H3K4 to *mPlrp2* and *mClps* promoters in the liver DNA of mice fed vitamin A-free diets (Fig.

5C, D).

A methylase inhibitor blocked hepatic *mPlrp2* and *mClps* expression and markedly lowered the levels of free retinol in the plasma and impaired hydrolysis of retinyl ester in the HSC-T6 cell line. AdoMet serves as a methyl donor in a series of biomethylation reactions, and the by-product, AdoHcy, is a potent feedback inhibitor of AdoMet-dependent methyltransferase. Therefore, the ratio of AdoMet to AdoHcy reflects the methylating capacity of tissues (27, 28). In order to clarify whether the changes of the ratio of AdoMet to AdoHcy are correlated with *mPlrp2* and *mClps* expression by the regulation of methylase, we examined the effects of a methylase inhibitor on *mPlrp2* and *mClps* gene expression in the liver. Methylthioadenosine (MTA), an effective methylation blocker, was used to treat mice fed the vitamin A free diet. Fig. 6A and 6B showed that severe attenuation of hepatic *mPlrp2* and *mClps* gene expression was observed in MTA-treated animals. Subsequent HPLC analysis showed that the peak of retinol from plasma extraction markedly declined in MTA-treated mice compared with control animals treated with saline. Quantitative analysis of peak levels revealed that MTA-treated mice exhibited significantly decreased levels of free retinol in the plasma compared with control animals (Fig. 6C, n=5). Fig. 6D and 6E indicated that MTA inhibited *mPlrp2* expression and hydrolysis of retinyl ester in a dose-dependent manner in the HSC-T6 cell line, and a linear correlation was found between the expression of *mPlrp2* and the levels of retinyl ester. These data indicated that *mPlrp2* and *mClps* are involved in the hydrolysis of retinyl esters in hepatic stellate cells in the liver and that their expression is regulated by DNA methylation.

Hepatic stellate cells specifically expressed *mClps* and *mPlrp2*. Hepatic stellate cells (HSCs) are primarily involved in the storage of retinol esters. Interestingly, cultured HSC-T6 cells in the activated state specifically expressed the *mClps* and *mPlrp2* genes (Fig. 7A), whereas other cell lines, including some hepatic cell lines, failed to exhibit detectable expression of *mClps* and *mPlrp2*. In order to examine the role of *mPlrp2* and *mClps* in the retinoid metabolism of HSCs, we performed gene silencing studies in the rat HSC-T6 cell line. As shown in Figure 7B and 7C, the knockdown of *mClps* by 80% in HSC-T6 cells led to the retention of more retinyl esters compared with control and scrambled shRNA treated cells, indicating a role for this gene in the hydrolysis of retinyl esters associated with the metabolism of retinoids in HSCs. Then we performed PLRP2 immunohistochemistry for establishing the intracellular site of *mPlrp2* expression in the liver, as showed supplementary Fig.2, *mPlrp2* specially expressed in portal tracts (rich hepatic stellate cells) in the liver.

Discussion

We are endeavoring to understand the basic biochemistry of retinoid metabolism, not only due to its fundamental scientific interest, but also for a better understanding of the role that vitamin A nutrition and retinoids play in health and disease. In normal nutrition conditions, the main site of vitamin A storage is the liver, where over 95% of total retinoids are found as retinyl esters (29, 30). The hydrolysis of retinyl esters is especially crucial in the digestion and intestinal absorption of dietary vitamin A and in the hepatic uptake, storage and mobilization of vitamin A. While PTL, PLRP2 and their partner, CLPS, were found to be the main hydrolases of retinyl esters in intestinal juice (19), the enzymes physiologically involved in hepatic retinyl ester hydrolysis remain unclear. Much early work showed that retinyl palmitate hydrolyzing activities are present in rat liver homogenates and that the activity varied markedly, with over a 50-fold range among the livers of individual rats (31), raising the question of whether these retinyl ester hydrolase activities detected in liver homogenates actually contributed to retinoids released in physiological livers. A few kinds of lipases are found in hepatocytes, such as hepatic lipase, carboxylesterases, and lipoprotein lipase. All of these known enzymes hydrolyze a variety of substrates, including retinoids, and no enzyme has been shown to be absolutely specific for retinyl esters (11).

In our previous work, a reduction in light illumination activated the expression of the pancreas specific lipases *mPlrp2* and *mClps* in the liver (20), and light-modulated retinoic acid signals regulated circulatory 5'-AMP levels by modulating CD73 activity, switching on *mPlrp2* and *mClps* gene expression in the liver (21). This study showed that temporary changes in the ratio of AdoMet to AdoHcy in specific tissues were associated with *mPlrp2* and *mClps* expression. That correlative relationship between this ratio and *mPlrp2* and *mClps* gene expression was supported by several findings: (i) a higher ratio of AdoMet to AdoHcy in the pancreas compared with the liver; (ii) an elevated hepatic ratio of AdoMet to AdoHcy in darkness or weak light illumination; and (iii) an increasing ratio of AdoMet to AdoHcy after 5'-AMP administration. In these independent studies on the elevation of the ratio of AdoMet to AdoHcy, the *mPlrp2* and *mClps* genes were found to be activated in liver tissues. This sensitive switch to both genes by temporary changes of the ratio of AdoMet to AdoHcy implied specific physiological functions. Homeostasis of free retinol in the plasma was controlled by vitamin A uptake from the diet and release of liver retinoid stores. A stable hydrolase activity in the degradation of liver retinoid stores was unreasonable for maintaining a relative constant circulatory level of retinol. The specific genes for hydrolyzing hepatic retinyl esters should be regulated by the status of the utilization of vitamin A in physiological conditions. Thus, both lipase genes *mPlrp2* and *mClps* could be optimal candidate genes involved in hepatic hydrolysis of retinol esters.

ATRA has been shown to play a negative role in retinol metabolism in the ferret liver (32).

Our current study demonstrates that in mice exposed to darkness, the ratio of AdoMet to AdoHcy decreased, inhibiting both *mPlrp2* and *mClps* gene expression in the liver, and that ATRA feedback inhibited hepatic *mPlrp2* and *mClps* gene expression by decreasing the ratio of AdoMet to AdoHcy. Moreover, mice fed a vitamin A-free diet also exhibited an elevated ratio of AdoMet to AdoHcy, strongly activating hepatic *mPlrp2* and *mClps* expression. We observed an increase in the binding of trimethylated H3K4 to the promoter sequences of *mPlrp2* and *mClps* in the liver of vitamin A-free mice. H3K4 trimethylation is a primary

signal that is sufficient for initiating a gene repression pathway *in vivo* (33). In vitamin A-free mice, increased H3K4 trimethylation relaxed this repression of related gene transcription. Moreover, a methylase inhibitor impaired both hepatic *mPlrp2* and *mClps* gene expression, resulting in the decrease of retinol levels in the plasma. Importantly, both of these genes were expressed specifically in an activated HSC-T6 cell line, and were only detected in portal tracts in the liver. Knockdown of these genes expression decreased retinol loss *in vitro*. Together, these observations strongly support the conclusion that hepatic *mPlrp2* and *mClps* were the primary genes involved in the hydrolysis of hepatic retinyl esters.

With the regulation of the light-modulated retinoic acid signal, liver tissues displayed a reverse alteration in the ratio of AdoMet to AdoHcy, which was an important metabolic indicator for cellular methylation status, also known as methylation potential (27). An altered ratio of AdoMet to AdoHcy resulted in protein carboxymethylation (34, 35), inhibiting or activating the expression of specific genes. In conclusion, the results of this study demonstrate that temporary changes in the ratio of AdoMet to AdoHcy switch hepatic *mPlrp2* and *mClps* gene expression, which are associated with the hydrolysis of hepatic retinyl esters for the utilization of vitamin A in the mouse liver.

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FOOTNOTES

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The abbreviations used are AdoMet, S-adenosylmethionine; AdoHcy, S-adenosylhomocysteine; mPlrp2, pancreatic related protein 2; mClps, procolipase; 5'-AMP, 5'-adenosine monophosphate; ATRA, all-trans retinoic acid; MTA, methylthioadenosine; PTL, pancreatic triglyceride lipase; REH, retinyl ester hydrolase; LED, light-emitting diode; IP, intraperitoneal; CD73, ecto-5'-nucleotidase; LD, light/dark; DD, dark/dark

Figure legends

Figure 1. Light intensity, but not specific wavelength, influenced expression of *mPlrp2* and *mClps* in the liver. (A) Specific expression of *mPlrp2* and *mClps* was detected in the pancreas in LD mice by RT-PCR, while no expression of both genes was found in the liver in LD mice. (B) Weak white light (<20 lux) induced *mPlrp2* and *mClps* gene expression in the liver in LD mice. The similar high intensity (about 250 lux) of white, red, yellow, green and blue light had the same inhibitory function on *mPlrp2* and *mClps* expression. DD mice were examined as a positive control. Results are representative of three independent experiments. *Gapdh* mRNA was monitored as an internal control.

Figure 2. Increased ratios of AdoMet and AdoHcy present in the pancreas compared with the liver under LD conditions. (A) Representative HPLC profiles of AdoMet and AdoHcy extracted from the pancreas. (B) HPLC profiles from the liver. Peak 1 is AdoHcy and peak 2 is AdoMet. The detection wavelength was 260 nm. (C) Quantification of AdoMet and AdoHcy in the pancreas and liver with respect to their internal protein concentration. Data are means \pm SEM, *p<0.05, n=6.

Figure 3. Darkness and weak light illumination elevated ratio of AdoMet and AdoHcy in livers of mice. (A) Representative HPLC profiles of AdoMet and AdoHcy extracted from mouse livers under LD, DD and light exposure in DD for 4 h. Peak 1, AdoHcy; Peak 2, AdoMet. (B) Quantification of AdoMet and AdoHcy in mouse livers under various light illuminations. Data are means \pm SEM, n=5, *p<0.05, compared to LD.

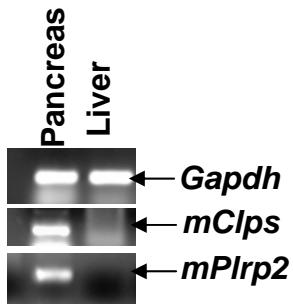
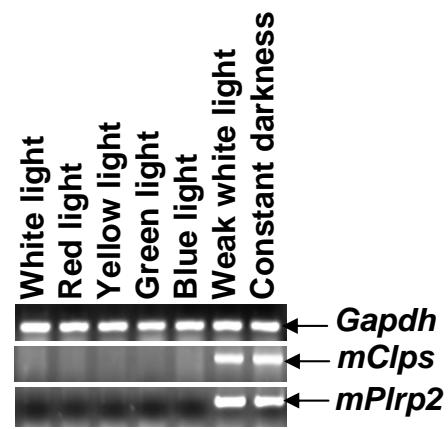
Figure 4. Effects of the administration of 5'-AMP and ATRA on the expression of the *mPlrp2* and *mClps* genes in the liver. (A) 5'-AMP (150 nmol/g) induced hepatic *mPlrp2* and *mClps* gene expression in LD mice, samples were collected 3.5 h after intraperitoneal injection. Results were expressed as double samples, representative of six animals in every group. *Gapdh* mRNA was monitored as an internal control. (B) Quantification of AdoMet and AdoHcy in livers of LD mice treated by 5'-AMP compared to mice treated with saline. An elevated ratio of AdoMet to AdoHcy was observed in mice treated by 5'-AMP. Data are means \pm SEM, n=6, *p<0.05. (C) Injection ATRA (2, 10 nmol/g) for 4 h inhibited hepatic *mPlrp2* and *mClps* gene expression in DD mice. Results are representative of six animals in every group. (D) Quantification of AdoMet and AdoHcy in the livers of DD mice treated with ATRA compared to mice treated with oil. Lowered ratios of AdoMet to AdoHcy were observed in mice treated by ATRA. Data are means \pm SEM, n=6, *p<0.05.

Figure 5. The *mPlrp2* and *mClps* genes are activated in the livers of mice fed a vitamin A-free diet. (A) RT-PCR analysis of *mPlrp2* and *mClps* mRNA in mice fed a vitamin A-free diet for 7 days. Results are expressed as double samples, representative of six animals in every group. *Gapdh* mRNA was monitored as an internal control. (B) Quantification of AdoMet and AdoHcy in the livers of mice fed a vitamin A-free diet compared to mice fed a normal diet. Increased ratios of AdoMet and AdoHcy were observed in vitamin A-free diet

mice. (C) Representative PCR profiles and (D) quantitative real-time RT-PCR analysis of the binding of histone H3K4-3Me to the *mPlrp2* and *mClps* promoter sequences in vitamin A-free and control mice, respectively. Data are means \pm SEM, n=6, *p<0.05.

Figure 6. MTA impairs the expression of *mPlrp2* and *mClps* in the livers of mice fed a vitamin A-free diet and hydrolysis of retinyl ester in the HSC-T6 cell line. (A) RT-PCR analysis of *mPlrp2* and *mClps* mRNA in vitamin A-free diet mice treated with MTA. Mice were placed on a vitamin A-free diet for 7 days and injected with MTA (100 nmol/g/day) on the last two days. Results were expressed as double samples, representative of six animals in every group. *Gapdh* mRNA was monitored as an internal control. (B) Quantitative real-time RT-PCR analysis of *mPlrp2* and *mClps* mRNA in vitamin A-free diet mice treated by MTA. (C) Quantification of retinol levels in plasma from extracted from vitamin A-free diet mice treated with MTA compared to mice treated with saline. Decreased plasma retinol levels were observed in MTA treated mice. (D) Quantitative real-time RT-PCR analysis of *mPlrp2* in HSC-T6 cells treated with the indicated dosages of MTA. (E) Quantification of retinyl esters from HSC-T6 cells treated with the indicated dosages of MTA compared to the control. HSC-T6 cells were incubated for 12 h with palmitic acid (100 μ M), after which the cells were treated with 5 μ M retinol for an additional 12 h, and then the indicated dosages of MTA were introduced into the cells. Retinyl ester concentrations were determined 12 h later. Data are means \pm SEM, n=6, *p<0.05.

Figure 7. The *mClps* gene is specifically expressed in the HSC-T6 cell line. (A) RT-PCR analysis of *mPlrp2* and *mClps* mRNA in multiple cell lines. (B) Total RNA from *mClps* knockdown HSCs was subjected to real time RT-PCR analysis. (C) Quantification of retinyl esters from knockdown HSCs compared to control and scrambled shRNA. HSC-T6 cells were incubated for 12 h with palmitic acid (100 μ M), after which the cells were treated with 5 μ M retinol for an additional 12 h, and then shRNAs were introduced into the cells. Retinyl ester concentrations were determined 12 h later. Results represent means \pm SEM from five HSC preparations; * P<0.05, ** P<0.01

A**B****Figure 1**

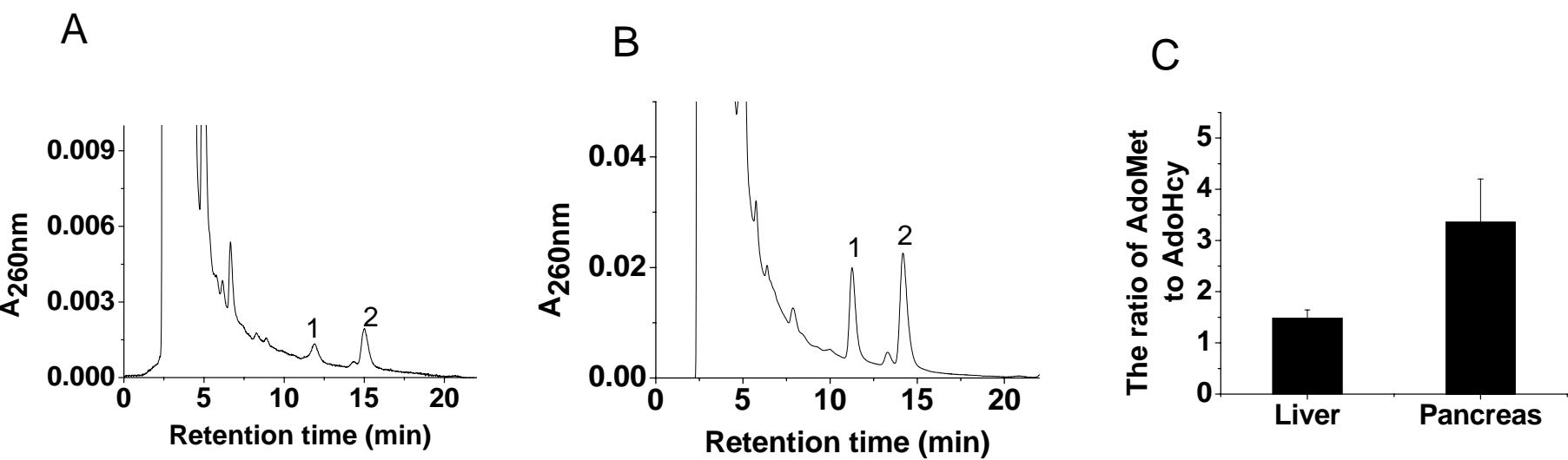


Figure 2

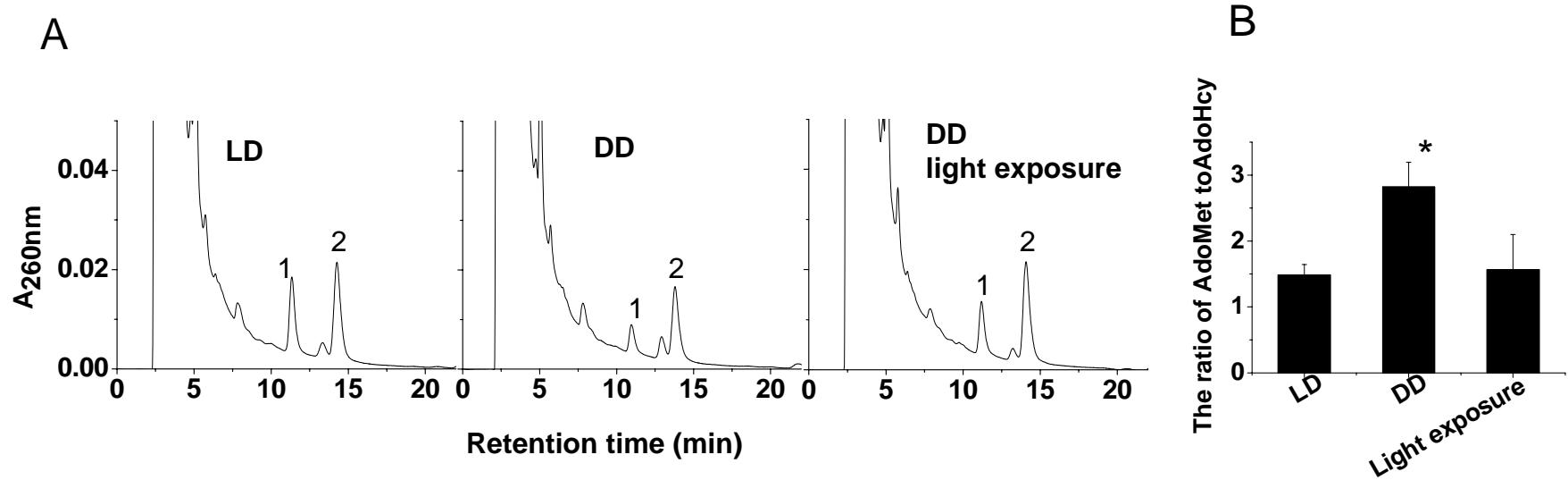
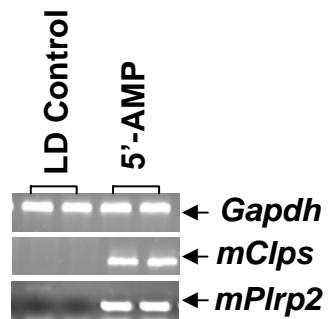
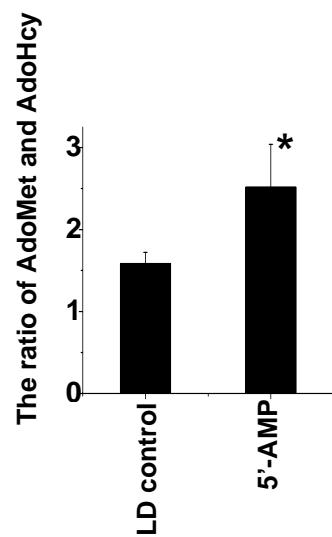
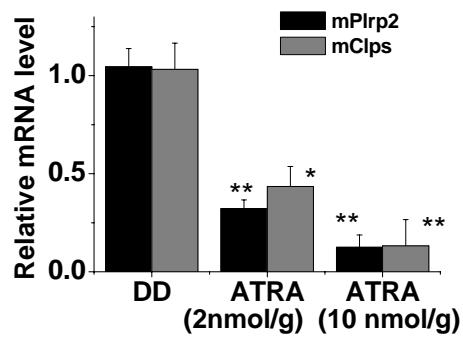
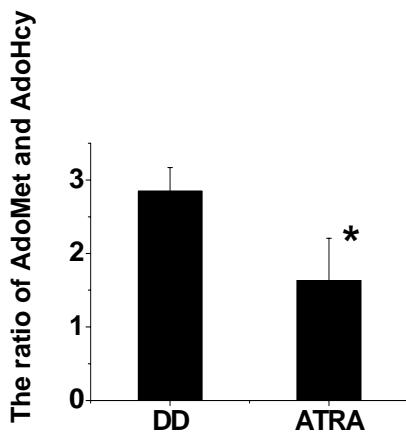
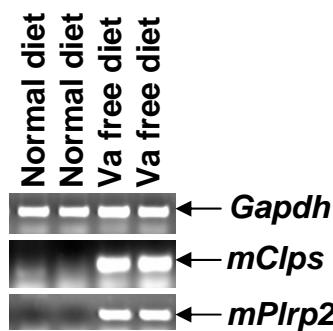
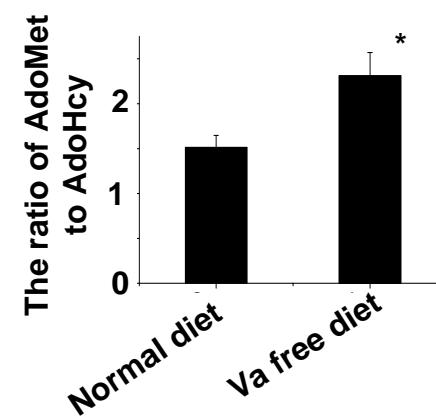
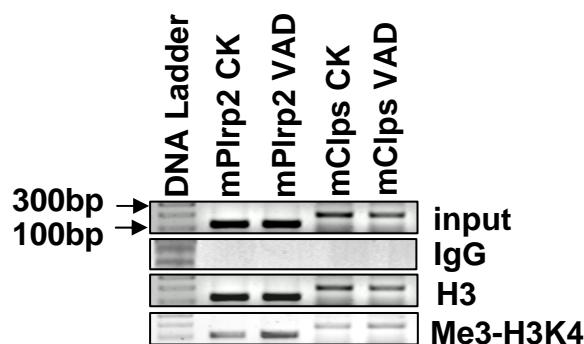
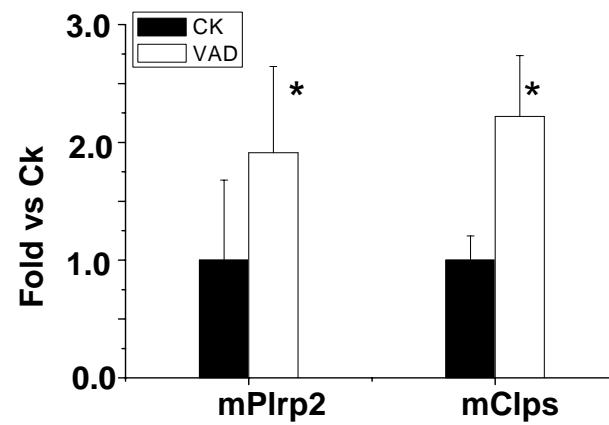
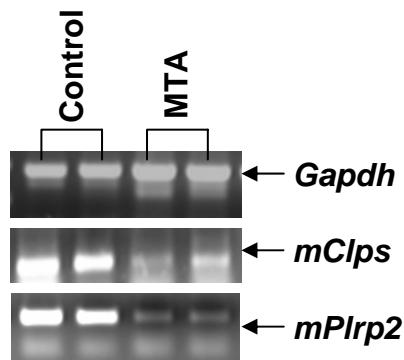


Figure 3

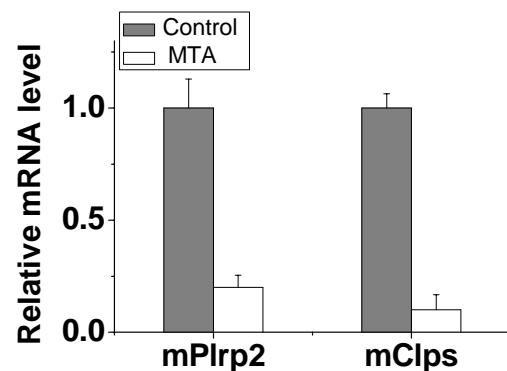
A**B****C****D****Figure 4**

A**B****C****D****Figure 5**

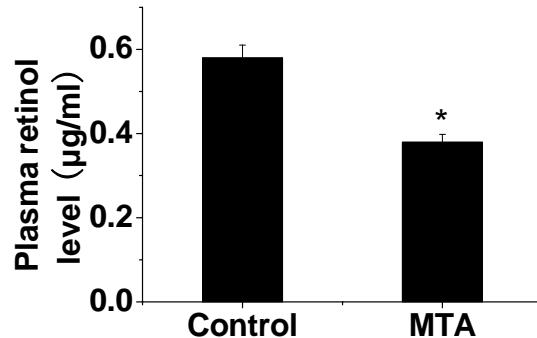
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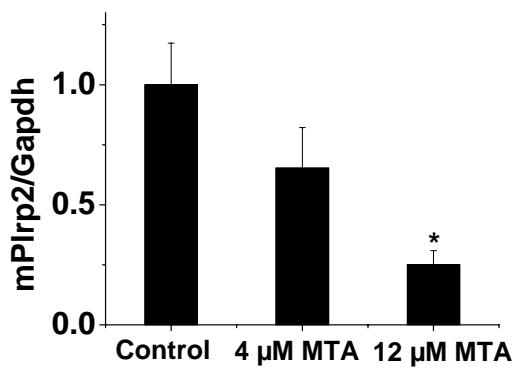
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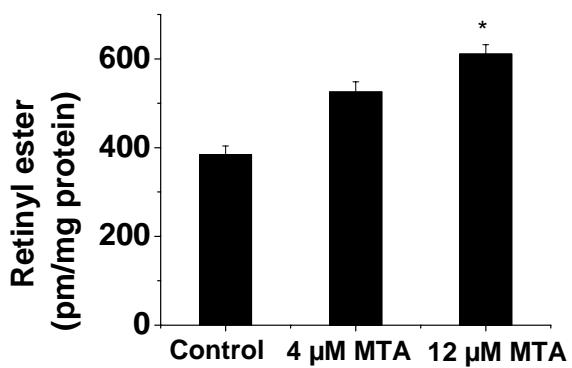
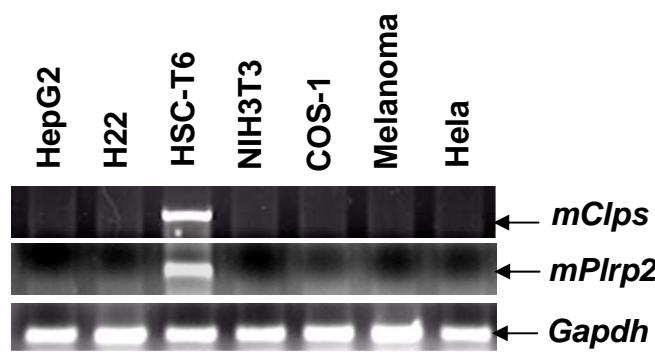
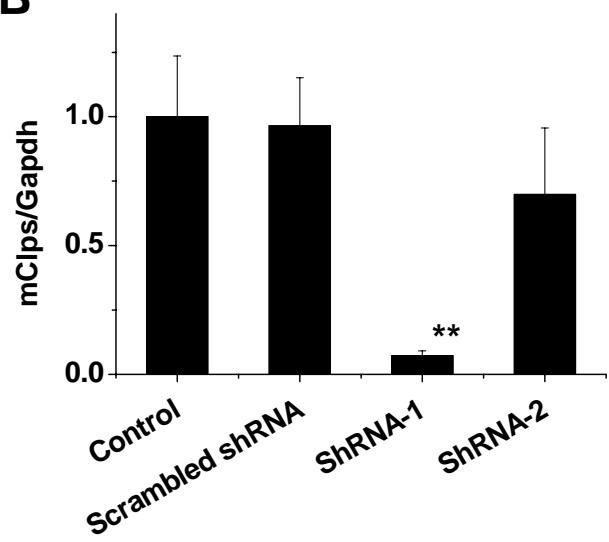
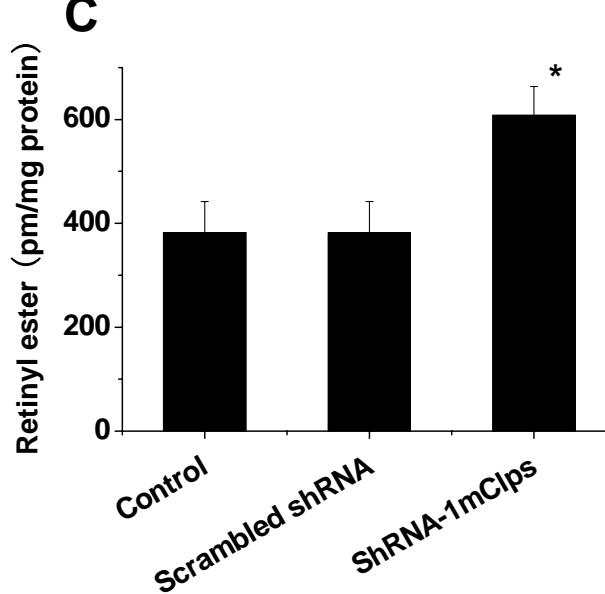


Figure 6

A**B****C****Figure 7**